

Virus Name: Kotonkan

Abbreviation: KOTV

Status: Probable Arbovirus

SALS Level: 2

Antigenic Group: rabies serogroup

Taxonomic status:

Other Information: None.

Select Agent:

SALS Basis: S

HEPA Filtration:

Section I - Full Virus Name and Prototype Number

Full Virus Name:

Kotonkan

Prototype Number:

IbAr 23380

Information from: G.E. Kemp and N. Karabatsos

Date:

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9/19/1984

Address: Division of Vector-Borne Viral Diseases, CDC, Fort Collins, Colorado 80522 USA

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Revised

Section II - Original Source

Isolated by: Dr. Vernon Lee

at: Ibadan, Nigeria

Genus and species: Culicoides spp., pool of approximately 2508 (1, 2) **Sentinel X**

Age/Stage: **Sex:**

Isolated From	Isolation detail
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Signs and symptoms of illness:

Arthropod engorged **depleted** **gravid**

Time held alive before inoculation: less than 8 hours

Collection date: 12/11/1967 **Method:** light trap

Place collected: University of Ibadan

Latitude: 7° 23' " N

Longitude: 3° 56' " E

Macrohabitat: grass pasture, derived from clearing high tropical forest

Microhabitat: cattle barns

Method of storage until inoculated: -60dF in mechanical freezer

Footnotes:

Section III - Method of Isolation and Validity

Inoculation Date: 12/14/1967

Animal: nb mice

Embryonated egg:

Tissue Culture:

(Details in Section VI - Biologic Char.)

Route inoculated: intracerebral

Reisolation: Not tried

Other reasons:

Homologous antibody formation by source animal (See Section II):

Test used: HI

CF

NT

Other:

Footnotes:

Section IV - Virus Properties

Physicochemical:

RNA:	DNA:	Single Strand:	Double Strand:
Pieces:	Infectivity:	Sedimentation coefficient(s): /strong>	
Percentage wt. of virion protein		, lipid	carbohydrate
Virion polypeptides:	Details:		
Number:	Details:		
Non-virion polypeptides:	Details:		
Number:	Details:		
Virion density:		Sedimentation coefficient:	
Nucleocapsid density		Sedimentation coefficient:	

Stability of infectivity (effects) pH

Lipid solvent:

(ether)	After treatment titer	Control titer
(chloroform) 10%	After treatment titer 2.5 dex	Control titer 4.4 dex

Detergent:

(deoxycholate)	After treatment titer	Control titer
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Other (formalin, radiation):

Virion morphology:

Shape: conical and parallel-sided, bullet-shaped virus

Dimensions: Mean (nm) 195x68 (2) range (nm) how measured thin-section electron microscopy (2)

Surface projections, envelope: surface projections; membranous envelope

Nucleocapsid dimensions, symmetry: cross-striations of nucleocapsid helix

Morphogenesis:

Site of constituent formation in cell: budding from plasma membrane (2, 3)

Site of virion assembly: viral matrix or inclusion material in cytoplasm (2, 3)

Inclusion bodies

Other

Hemagglutination:

Hemagglutination	Not tried	Antigen source	
Erythrocytes	pH range		pH optimum
Temperature optimum		range	

Remarks

Serologic methods recommended: CF, serum dilution NT in nb mice * Mixed pool c

Footnotes:

Section V - Antigenic Relationship And Lack of Relationship To Other Viruses

In CF tests done at Ibadan, a kotonkan virus antigen reacted with immune mouse ascitic fluid for Mokola virus of the rabies serogroup but not with nine arbovirus multivalent sera nor with monovalent immune fluids prepared with 64 distinct agents, including rabies, Lagos bat, Chandipura, bluetongue, Ibadan isolates of the epizootic hemorrhagic disease of deer group, IbAr 23388 (Abadina), herpesvirus, Newcastle disease and vaccinia. Following the isolation and identification of bovine ephemeral fever (BEF) virus in Nigeria, kotonkan virus was tested for CF reactions with this agent. Kotonkan hyperimmune mouse ascitic fluid with homologous titer of 1:32 failed to react at the 1:4 dilution with BEF mouse brain antigen; BEF hyperimmune mouse ascitic fluid (homologous titer 1:64) did not react at the 1:4 dilution with kotonkan antigen [2].

In cross-CF tests done at the Yale Arbovirus Research Unit, kotonkan virus again reacted with Mokola but not with rabies, Lagos bat, Obodhiang or 191 other viruses. Indirect FA tests, done at the Centers for Disease Control, confirmed the CF relationship between kotonkan and Mokola [2], [3]. In N tests, however, kotonkan virus failed to react with Mokola and other rhabdoviruses [2].

Virus or Antigen	Cross-complement-fixation tests with kotonkan and rabies serogroup viruses [2]					Neutralization tests with kotonkan and other rhabdoviruses [2]						
	Mouse ascitic fluid CF antibody to:					Mouse ascitic fluid NT antibody to:						
	Kotonkan	Mokola	Rabies CVS	Lagos bat	Obodhiang	Kotonkan	Mokola	Rabies ^b CVS	Lagos bat	CHP	BEF Afr ^c / Aust ^c	
Kotonkan	64 ^{**}	32	0	0	0	2.9 ^a	0.2	1.0	0.6	0.4	0.4	0.4
Mokola	0	256	32	4	16	0.0	3.1	0.9	2.6			
Rabies, CVS	0	64	256	0	0	0.0	0.2	3.0	0.2			
Lagos Bat	0	128	16	32	8	0.0	0.1	0.1	2.0			
Obodhiang	0	4	0	0	128							
191 arbo- viruses and other viruses of vertebrate	0	0	0	0	0							

^{**} ascitic fluid titer; 0 = <4

^a LNI expressed in dex

^b Horse serum supplied by Lederle Labs.

^c Supplied by Veterinary Res. Lab., Onderstepoort, South Africa.

Section VI - Biologic Characteristics

Virus source (all VERTEBRATE isolates):

Lab Methods of Virus Recovery (ALL ISOLATIONS): Newborn mice

Susceptibility of Cell Culture Systems:

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Aedes albopictus (CL)	MB 9							+ (4)
Vero (CL)	MB 9 AA 1	7-10	3+(5)		6	2mm (4)		
BHK-21 (CL)		7-10	3+(4)					
CER (CL)	CER 4			4.0-5.0^d (6)				
BHK-21 (CL)								- (9)
Vero (CL)		4-5	CPE	5.8 (9)				
E6 (CL)		5-6	+ -CPE	4.5 (9)				
C6/36 (CL)		3	No CPE	3.8 (9)				+ (9)

NOTE: Original Vero or BHK-21 cell culture passage was co-cultivated with infected Ae albopictus cells (4, 5).

D Expressed in dex

Section VII - Natural Host Range

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Culicoides spp. *	1/72 (14,500 insects)		University dairy barn, Ibadan, Nigeria (1, 2) Northern Nigeria (2)
Man		2/39 NT	Jos
Trade cattle		124/128 NT	(Northern origin)
Dairy cattle		29/29 NT	Vom
Horses		0/2 NT	
Cattle egret, Bubulcus ibis		1/1 NT	Bassa
Hedgehog, Atelerix albiventris		14/15 NT	
Giant rat, Cricetomys gambianus		14/21 NT	
Other rodents		1/2 NT	
Other rodents		0/2 NT	Fika Southern Nigeria (2)
Man		0/87 NT	South Nigeria
Monkey		0/1 NT	University of Ibadan
Swine		0/50 NT	
Sheep		8/75 NT	
Cattle		24/25 NT	
Chickens		0/16 NT	

Horses	2/33 NT	Ibadan
Wild birds	0/41 NT	
Shrew, Crocidura sp.	0/48 NT	University of Ibadan
Rodents	0/43 NT	
Bats	0/23 NT	

* In typical light trap catches during this period, the most abundant species was *C. pallidipennis*.

Section VIII - Susceptibility To Experimental Infection (Record Viremia)

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log ₁₀ /ml
mice (nb)	IbAr 23380, MB 0	ic	1/8 dead, 1/8 sick (2)	11	
mice (nb)		ip			
mice (nb)		sc			
mice (wn)		ic			
mice (wn)		ip			
mice (nb)	IbAr 23380, MB 9	ic	illness, death (2)	14	
Fulani calves, white (3 mo)	IbAr 23380, MB 3	sc, intradermal	1/2 developed mild clinical illness similar in many respects to that caused by bovine ephemeral fever virus. Viremia not detected during 15 days post-inoculation. Both calves developed NT antibody following infection (7).		

